The Asymmetric Synthesis of α -Amino and α -Hydrazino Acid Derivatives via The Stereoselective Amination of Chiral Enolates with Azodicarboxylate Esters.

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Abstract: The utility of azodicarboxylate enters as (+)NH₂ and (+)NH-NH₂ synthons in highly diastereoselective reactions with chiral carboximide-derived enclases has been demonstrated. The lithium enclates derived from 4-substituted N-acyl 2-oxazolidinones were found to react with di-tert-butyl azodicarboxylate (DBAD) to afford the derived 2-hydrazido carboxylic acid derivatives in yields in excess of 90 %. The diastereoselectivities of these reactions ranged from 97% to greater than 99%. The subsequent transformation of these adducts to both α-hydrazino and α-amino acids in enantiomeric purities in excess of 99% is described.

Introduction

Although the exceptional electrophilic reactivity of azodicarboxylate esters has been widely recognized, 1,2,3,4,5 the application of this reagent to the electrophilic amination of carbon nucleophiles has remained largely undeveloped. This is somewhat surprising since the first documented example of this process, the reaction of diethyl malonate with diethyl azodicarboxylate (DEAD), appeared in 1924 (eq 1).6 The subsequent applications of this reagent as an (+)NH2 synthon has been limited to a few reports concerning the reaction of DEAD with electron-rich aromatics and enol ethers (eq 2).8 At the time that this study was initiated, the single example illustrating the use of an organometallic reagent as a carbon nucleophile in this context was the reaction of tert-butylmagnesium chloride with di-tert-butyl azodicarboxylate (DBAD).9,10

The above precedents suggested that axodicarboxylates possessed a desirable combination of high reactivity toward nucleophiles coupled with good shelf stability. Accordingly, the present investigation describes the reactions of axodicarboxylate esters with the illustrated chiral enolate system (eq 3,4) as a general route to both α -hydrazino and α -amino acids. 11,12 Currently available methods for obtaining α -hydrazino acids in enantiomerically pure form are limited either to classical resolution, 13 or to multistep procedures starting from the requisite enantiomerically pure α -amino acid precursors. 14,15,16 A convenient, general method for the

asymmetric synthesis of these compounds based on the illustrated reaction of chiral carboximide enolates with a suitable azodicarboxylate electrophile would have obvious value, and could greatly facilitate the development of the biological potential offered by this class of compounds.

In preliminary studies, the suitability of azodicarboxylates as (+)NH2 synthons toward carboximide enolates was established. Our selection of Di-tert-butyl azodicarboxylate (DBAD) (1) as the reagent of choice was motivated by several factors. First, it is a stable, crystalline yellow solid (mp 90-92 °C) that is readily prepared, and is also commercially available. 17 Second, since the tert-butoxycarbonyl (Boc) group is widely used as a protecting group in peptide chemistry, methods for its removal under mild, nonracemizing conditions were well established, and are complementary to known methods for N-N bond cleavage. This offered the potential for convenient access to either \alpha-amino or \alpha-hydrazino acids from a common intermediate. Finally, preliminary data suggested that this reagent possessed a unique combination of high reactivity and high diastereoface selection in reaction with chiral imide enolates.

Results and Discussion

Electrophilic Amination of Chiral Imide Enolates. The carboximide substrates 3a-h chosen for this study were prepared as previously described via re-

action of the lithiated oxazolidone 2 and either the desired acid chloride 18 or the analogous mixed pivalic acid anhydride, which was generated in situ (eq 5). Initial electrophilic amination studies were conducted on the dihydrocinnamate derivative 3c (R=CH2Ph) which was chosen as a representative substrate. The reaction of its derived lithium enolate, generated with lithium diisopropylamide (LDA) (1.05 equiv) in THF as previously described, 19 with DBAD (1.2 equiv) at -78 °C was found to be instantaneous, as evidenced by the immediate decoloration of the cooled (-78 °C) solution of DBAD in CH2Cl2 upon its addition to the esolate solution by rapid cannulation. Following an almost immediate quench (1 min) with glacial acetic acid (2:6 equiv) and conventional isolation, the adduct 4c (R=CH2Ph) was obtained in excellent yield (eq 6).

The ¹H NMR spectrum (CDCl₃, 300 MHz) of 4c obtained at 23 °C was largely uninterpretable due to hindered rotation about the Boc groups; however, the corresponding spectrum obtained at 57 °C was dramatically simplified and readily analyzed. This analytical method revealed only one product diastereomer; however, significant broadening of the spectral lines even at this temperature made it impossible to verify the presence of small but significant quantities of the minor diastereomeric impurity. In order to accurately determine the diastereoselectivity in these reactions, an unambiguous method for the production of both diastereomers was developed. One such method which proved to be viable is outlined in Scheme 1.

Scheme 1

The lithium enolate of the corresponding benzyl ester 5 was treated with DBAD in a similar fashion, to afford the racemic α-hydrazido ester 6c in 60% yield. Following benzyl ester hydrogenolysis, formation of the mixed pivalic acid anhydride, and subsequent treatment with the lithiated oxazolidone (2.1 equiv), the desired carbox-

imide 4c was obtained in 91% overall yield (from 6c) as a 1:1-mixture of C_2 diastereomers. These compounds proved to be readily separable by both TLC as well as by HPLC (α =1.66). The faster-eluting component of this diastereomeric mixture was found to correspond to the major diastereomer from the imide-DBAD reaction, and was subsequently shown to possess the 2(S) configuration by correlation with L-phenylalanine (vide infra). HPLC analysis of the unfractionated imide-derived sample of 4a afforded a 2(S):2(R) ratio of 97:3, which was increased to >200:1 (91% yield) on a ten-gram scale following a single chromatographic purification on a standard grade of silica.

In order to facilitate the diastereomer analysis for other enolate-DBAD reactions, a more convenient method for obtaining hydrazido imide samples enriched in the minor diastereomer was developed. It was subsequently discovered that base-catalyzed epimerization of 4 could be achieved under the proper conditions. For example, when a solution of the 2(S) diastereomer of hydrazido imide 4c (0.09 M in THF) and 2 equiv of 1,1,3,3tetramethylguanidine was heated at reflux for 4.5 h, a 45:55 mixture of the 2(S) and 2(R) diastereomers respectively was obtained in 63% yield. With appropriate adjustments for differing substrate epimerization rates,20 this method was generally employed to independently verify the identity of the minor 2(R) diastereomer in those cases where sufficient quantities could not be isolated directly from the amination reaction for spectral characterization.

The results obtained for the reaction of DBAD with the lithium enclates derived from imides 3a-f under reaction conditions similar to that employed above are listed in Table 1 below.

Table 1. Selective "Amination" of Carboximide Enclates (eq 6).

Entry	Imide	Substituent (R)	Kinetic Ratio ⁴ 2(S): 2(R)	Product	Yleki, ^b %
A	34	Me	98 : 2 ⁰	40	92
8	36	ан,ан-ан,	98:2	4b	94
С	3 c	CH ₂ Ph	97:3	4 ¢	91
D	3d	Ph	97 : 3	4d	96
ε	30	CHMe ₂	98:2	40	95
F	31	CMe ₃	>00:1	#	96

Ratios determined by HPLC analysis. ^bValues rater to tsolated yields of pure adduct, 2(8): 2(R)> 200:1. ⁵eolated yield of the dissersomer mixture.

Minor modifications of the above chromatographic procedure afforded diastereomerically pure hydrazides 4b-f [2(S):2(R)>200:1] in 91-96% yields. The chiral auxiliary proved to be not only an effective directing ligand in the amination reaction, but it also performed as an efficient chiral chromatographic resolving moiety as well. In accord

with previous studies, a stereoregular elusion order was observed with the illustrated series of hydraxides, with the major 2(S) diastereomer exhibiting the greater mobility (vide infra).

In addition to the cases summarized in the Table, the direct amination of several less conventional substrates was also examined (eq 7, 8). The glutaryl imide 3g was selectively deprotonated with 1.0 equiv of LDA (-78 °C, 5 min), and the resulting solution was treated with a slight excess of DBAD at -78 °C for 40 s prior to the usual acetic acid quench. Following a conventional isolation and chromatographic purification, the amination product 4g was obtained in 51% yield (diastereomeric purity ≥95%) along with a 16% recovery of the imide starting material 3g.21 This is the only case that we have encountered in which the product diastereomers did not afford a baseline separation on analytical HPLC. In this instance, the reaction diastereoselection was conservatively estimated to be ≥95:5 from the 300 MHz ¹H NMR (331 °K) spectrum of the unfractionated product.

The lithium dienolate of the β , β -dimethylacryloyl imide 3h was similarly treated with DBAD to afford a 51% of the 1,2-adduct 4h along with 42% of the 1,4-addition product 7 as a 6:4 mixture of E:Z isomers (eq 8). Again, the 1,2-adduct was formed with high stereoselectivity (98:2), and the major product could be isolated in >200:1 diastereomeric purity (48% yield) on a multigram scale after chromatographic purification.

The amination reaction of these carboximide lithium enolates with DBAD show striking similarities, as well as significant differences, to analogous imide enolate-electrophile reactions that have been investigated in these laboratories. As in the analogous alkylation, ²⁴ acylation, ²⁵ hydroxylation, ²¹ halogenation, ²³ and azidation, ²⁴ studies, the sense of asymmetric induction observed in this study is consistent with preferential attack of DBAD on the least hindered diastereoface of the chelated (2) enolate.

Of all of the above imide enclate-electrophile reactions investigated, these amination reactions have simultaneously proven to be among the fastest, highest yielding, and most uniformly stereoselective of any yet encountered. The broad scope of this reaction is obvious from the data in Table 1. Unlike the analogous alkylation reactions, the reactivity of DBAD toward these enclates was not perceptibly attenuated as the steric requirements of the enolate substituent (R) are increased. In fact, not only was a high level of reactivity maintained, but the stereoselectivity was significantly improved. A striking illustration of these facts is provided in Entry F, in which the tert-butylacetyl derivative 4f was obtained in 96% isolated yield, with >200:1 kinetic stereoselection, after a 3-min reaction time at -78 °C. In studies of the reactions of these chiral imide enolates with other heteroatomic electrophiles, it has been generally observed that the kinetic stereoselectivity is amplified with increasing steric requirements of the enolate R group. However this dependence of the reaction stereoselection on the structure of the imide enolate acyl residue is significantly attenuated in the corresponding reactions with DBAD. These results are in direct contrast to analogous halogenation,23 hydroxylation,²¹ and azidation studies,²⁴ in which a significant decrease in reaction stereoselection has been observed for phenylacetate-derived imide substrates (e.g. 3, R = Ph) as compared to the corresponding propionate (R = Me) and butyrate (R = Et) derivatives.

Mechanistic Considerations. We speculate that these highly exothermic reactions proceed through an early reactant-like, but highly ordered, transition state. Since DBAD can be considered a nitrogen analog of a conventional ox, \beta-unsaturated ester, e.g. di-tert-butyl fumarate, these reactions probably bear a strong mechanistic relationship to the Michael addition reactions of enolates to such esters, a reaction which is the focal point of some current interest. Several models for this reaction have recently been extended to explain the diastereoselectivity observed in acyclic systems. Studies have recently appeared in support of a pericyclic process, 25 in which the enone carbonyl is coordinated to the enolate metal while an alternative open transition structure, involving no metal ion organization, has also been proposed.²⁶ Although much of the data that has been accumulated can be rationalized with either model, cases have been reported that are most readily explained in terms of a pericyclic transition state. In accord with the above precedent, we favor an analogous pericyclic transition state for the reaction of DBAD with imide enolates. Several possible transition structures for this reaction are illustrated below. It is evident that a high degree of organization can be achieved through coordination of the lithium atom with either the ester carbonyl (structure T1) or the azo nitrogen of DBAD (structures T2 and T3).

In the former case, an 8-centered pericyclic transition state is thereby involved, while in the laster cases, 6-centered pericyclic structures which are directly analogous to the now commonly accepted Zimmerman-Traxler aldol transition state²⁷ appear plausible. In these laster two cases, al-

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though DBAD could, in principle, assume two possible orientations as illustrated in T_2 and T_3 , it appears that the former transition structure T_2 might be favored on steric grounds.

The chromatographic parameters for the α -hydrazido carboximide diastereomers prepared during this study are summarized in Table 2. As in the examples cited above, the major 2(S) product diastereomer is seen to elius first in each case. The separation factors (α) were found to be large enough so that diastereomer resolution could be effected on silica gel with little difficulty. The possible ground state conformations which may be employed to rationalize the stereoregular chromatographic behavior displayed by the diastereomeric α -hydrazido carboximide adducts are illustrated below. It is suggested that the major, faster-eluting diastereomer 2(S)-4 adopts the illustrated conformation in which the lipophilic moieties, R and Bn, "shield" both faces of the molecule from the polar adsorbant.

It is felt that the minor diastereomer 2(R)-4, on the other hand, might adopt the corresponding conformation in which these substituents reside on the same molecular face. Since the opposite, more polar face is exposed to the adsorbant, this diastereomer is more strongly retained. In each case, conformational organization is achieved by a combination of both intramolecular hydrogen bonding and the preferential opposition of the carbonyl dipoles.²⁸

Table 2. HPLC Resolution of a-Hydrazidoacyi Diastereomers 4.*

Imide	Substituent (R)	k ^{. b} 2(S)-4	k ^{-b} 2(R)-4	Sep. Factor ((x)
42	Me	2.35	3.27	1,39
4b	сн,сн-сн,	2.97	4.64	1.58
4 c	CH _g Ph	3.84	6.37	1.66
4d	Ph	2.88 ^d	4.97	1.73 ^d
40	CHMe ₂	2.95	4.53	1.54
48	CMe ₃	2.65	4.29	1.62
4h	C(Me)C=CH _g	3.02	4.14	1.37

^{*}Analyses carried out on a Waters 5 micron Radial-Pak column eluting with CH₂Cl₂-hazane-MeCN (70:30:5). HPLC capacity factor defined as (t - t₀) A₀, * HPLC separation factors for product disstancement defined as the ratio of the capacity factors.

Eluting solvent: CH₂Cl₂-hexane-MeCN (70:30:7.5).

This "adsorption" model may also be applied to the α -hydroxy carboximides, 21 and it also predicts the elution

order for other diastereomeric oxazolidones as well.²⁹

Imide Hydrolysis and Transesterification. With a general method for the preparation of enantiomerically pure a-hydrazido carboximides in hand, attention was directed to the development of methods appropriate to the mild hydrolysis and alcoholysis of the hydrazide adducts (eq 9). At the outset, it was unclear how difficult this would be to achieve for a broad range of substrates. Although the exocyclic imide carbonyl is electronically activated by the ct-heteroatom substituent, in the present case that substituent is also sterically demanding. From previous studies in these laboratories, it was known that the reactivity of the exocyclic imide carbonyl moiety toward nucleophilic attack is suppressed with increasing steric demands of the exocyclic acyl substituent. In such instances, competing attack of the nucleophile at the endocyclic auxiliary carbonyl function can dominate, resulting in the preferential formation of oxazolidone ring cleavage products (eq 10).

In order to compare the effectiveness of several reagents that had previously been used to effect carboximide hydrolysis or transesterification, four prototype hydrazido imides were studied. Hydrazide 4c (R=CH2Ph) was chosen to represent a conventional substrate, while the isovalerate (4e, R=CHMe2) and tertbutylacetate (4f, R=CMe3) derivatives were chosen to test the steric limitations regarding the positional selectivity of these reagents. Finally, the phenylacetate derivative (4d, R=Ph) provided a sensitive test of the potential for racemization which might occur during these processes.

For this study, only enantiomerically pure (2S:2R > 200:1) imide samples were employed so that even minor amounts of racemization could be detected with high precision by subsequent capillary gas chromatographic analysis of the derived a-amino methyl ester (+)-MTA amides (vide infra). The results of this study reveal several interesting trends (Table 3). For the conventional substrate 4c (Entries A-C), all three methods examined, hydrolysis, methanolysis, and benzyl ester transesterification, 19 provided the acid (ester) derivative, 8c, 9c, and 6c, respectively, in good to excellent yield with no detectable racemization, along with a corresponding recovery of the chiral auxiliary. However, varying amounts of racemization were observed for the phenylacetate derivative 4d depending upon the reagent employed (Entries D-F). The preferred method for removing the chiral auxiliary in this case proved to be LiOH hydrolysis, which provided the corresponding acid in good yield (84%) with minimal (\$2%) racemization. On the other hand, magnesium methoxide methanolysis afforded the corresponding methyl ester 9d in 93% ac, while the benzyl ester 6d obtained following treatment with LiOBn was almost totally racemic. A comparison of Basries G and H, and Entries I and I reveals the substantially improved exocyclic carbonyl positional selectivity displayed by LiOBa as compared to both LiOH and BrMsOMe for imide deacylation reactions when the steric interactions in the substrate become more severe. These results are in accord with prior precedent established in this laboratory. Although the yield of the isovalerate benzyl ester 6e obtained with LiOBn was still very good (82%) (Entry H), the tertbutylacetate derivative 4f (Entry J), suffers from an elevation of the undesired mode of cleavage (eq 10) resulting in significantly reduced yield (51%) of the desired product 61. However, even in this extreme case, excellent exocyclic carbonyl cleavage regioselectivity could be restored with LiOOH, which afforded the desired acid 8f in 91% yield (>99% ee) along with a 91% recovered yield of the oxazolidone chiral auxiliary 2 (Entry K). It should be noted that in none of the transesterification and hydrolysis experiments conducted on the sterically hindered substrates 4e and 4f was any racemization detected (Entries H-K).

Table 3. Hydrolysis & Transesterification of Hydrazide Adducts (eq 9).

Entry	Compnd.	Reegent	Conditions *	Product	Yield, %	oo, %°
Ā	4c R-8n	ПОН	0 °C; 3 h	le*	824	> 99
В	4c	BrMgOMe	0 ℃; 0.5 h	Sc	80	> 90
C	4 c	UOBn .	0 ℃; 2 h	€ c	96	> 99
D	4d R-Ph	UOH	0 °C; 2 h	844	84	98
E	4d	BrMgOMe	0 ℃; 1.6 h	9d	71	93
F	44	UOBn	0 °C; 4.8 h	84	80	22
G	4e R.→ Pr	BrMgOMe	0 °C; 22 h	80	12	
н	40	UO B n	0 ℃; 15 5 h	60	82	> 90
1	44 R=4-Bu	LIOH	0 ℃; 16 h	W 4	16	> 90
J	4	UO B n	0 °C; 60 h	•	61	> 99
K	4	ПООН	0 °C; 3.2 h	M.	91	> 90

*UOH hydrolyses conducted in 2:1 THF-HOH with 2.3 equiv of UOH (substrate cond. 0.17 M). Methanolyses were conducted with 2.0 equiv of McOMgBr in McOH (substrate cond. 0.04 M). Benzyl ester transesserifications were conducted with 2.0 equiv of UOBn in THF (substrate cond. 0.14 M). Percade-mediesel hydrolyses were performed with 2.0 equiv of UOH/ 4.0 equiv of HOOH in 3:1 THF-HOH (substrate cond. 0.05 M). ** Yields quoted are for the overall conversion to 8. ** The se values were determined by capillary GLC of the derived MTPA amides. ** Carbonylic acide 8 were characterized as their methyl esters 9.

The unparalleled reactivity and exocyclic carbonyl regioselectivity displayed by LiOOH in carboximide hydrolysis (Entry K) has been found to be quite general, and as a consequence, it has proven to be the reagent of choice for removal of these chiral auxiliaries from sterically demanding carboximide substrates. These results have been discussed in some detail elsewhere.³⁰

a-Amino Acid Synthesis and Optical

Purity Assay. Having established that these chiral imide enclates could be employed in a general route for the preparation of enantiomerically pure N-protected ahydrazino esters and ecids, a practical protocol for the conversion of these substrates to ox-amino acid derivatives was developed. Ideally the preferred mode of degradation might involve the direct N-N bond cleavage of the urethane-protected hydrazine to afford the N-Boc a-amino acids (esters), since these products would be convenient and versatile, N-protected α-amino acid synthous. However, in earlier studies conducted in these laboratories, attempts to effect this transformation on the diacyl hydrazides via Raney nickel hydrogenolysis, or dissolving-metal reduction proved unsuccessful.31 However, an alternative method consisting of prior removal of the Boc groups followed by hydrogenolysis of the resulting hydrazino esters over Raney nickel did provide a useful route to the desired α -amino products.³² The overall process was combined with an acylation step with (+) MTPA chloride³³ (eq 11). Diastereomer analysis of the resulting (+)-MTPA amide derivatives 10 by capillary GLC provided a sensitive assay of the stereochemical fidelity of the entire sequence (vide infra). The representative substrates were first converted to the corresponding methyl esters 9. In the case of the benzyl esters 6, this was achieved in quantitative yield via hydrogenolysis over 5% Pd-C (1 atm H₂, EtOAc, 25 °C) followed by diazomethane treatment.

The remaining steps in the sequence are illustrated by the following representative example (Table 4, Entry A). Hydrazido ester &c (R=CH₂Ph) was deprotected to the free hydrazine and the resulting solution was directly hydrogenated over Raney nickel catalyst (500 psi of H₂, 4 h, 25 °C).

Table 4. Reduction and Acytetion of α-Hydrazido Estere (eq 11).

Entry	Hydrazide	(R)	H _e time (h)	Product	Yleid %	Resto * 2(S) : 2(R)
A	6 c	CHLPh	4	10c	94	>200 : 1
8	9d	Ph	4	10d	99	90:1
C	60	CHMe ₂	16	10e	83	>200 : 1
D	•	CMe,	16	107	89	>200 : 1

^{*} Determined by capitlary GLC of the product(+)-MTPA amides 10.

The unpurified α -amino ester, obtained after filtration through Celite and solvent removal, was acylated with (+)-MTPA chloride to afford the (+)-MTPA amide 10c in 94% overall yield for the three steps. This material was found to be identical to the (+)-MTPA amide derived from authentic L-phenylalanine, thereby establishing the 2(S) absolute configuration in the synthetic phenylalanine intermediate. Analysis by capillary GLC revealed the ratio of 2(S):2(R) diastereomers to be >200:1. The additional

examples provided in Table 4 were subjected to the identical reaction sequence, differing only in the longer Raney nickel hydrogenolysis time (16 h) employed for the hindered substrates 6e, and 6f (Entries C and D). From the tabulated data, it is apparent that a wide variety of camino acids can be obtained in this fashion in good yield with insignificant racemization.³⁴

In conclusion, the electrophilic amination of chiral imide enolates with DBAD provides an expedient approach to the asymmetric synthesis of both α -hydrazino and α -amino acid derivatives. A wide range of target structures of predictable absolute configuration are accessible in high enantiomeric purity, and excellent overall

yield. This includes classes of α -amino and hydrazino acid derivatives such as aryl and tert-alkyl glycines and their N-amino congeners which are not readily accessible by complementary methods. This approach to the electrophilic amination of carboxylic acid enolate synthons is highly complementary, although not quite as efficient, as the electrophilic azidation of these same enolate systems recently reported from this laboratory. An in view of the efficiency with which the azidation process may now be carried out, the present methodology should be principally exploited within the context of asymmetric α -hydrazino acid synthesis.

Experimental Section

General. Di-tert-butyl azo-dicarboxylate (1) (DBAD) was purchased from Pluka AG and used as received. The imides 3 were prepared as previously described from the corresponding acid chlorides or mixed pivalic acid anhydrides and (45)-4-Phenylmethyl-2-oxazolidinone (2) (XpH).³⁵ Lishium disopropylamide (LDA) was generated in situ by treating dry diisopropylamine (0.38 M in THF) with 0.92 equiv of n-butyllithium (1.6 M in bexane) at -78 °C for 30 min. High pressure Raney Ni hydrogenolyses were performed in a magnetically stirred, glass-lined, stainless-steel autoclave. Flash chromatography was performed on E. Merck silica gel 60 (230-400 mesh). Solvent gradients for medium pressure preparative chromatography (MPLC) were constructed with a simple two-chamber apparatus³⁶ which supplied, by gravity feed, a single high-capacity pump. Michel-Miller columns dry-packed with 230-400 mesh silica gel were used in conjunction with this apparatus.

(4S)-3-(1-oxo-4-pentenyl)-4-(phenylmethyl)-2-oxazolidinone $(3b, R = CH_2CH = CH_2)$. To a mechanically stirred solution of 5.61 mL (5.61 g, 55.0 mmol, 1.1 equiv) of 4-pentenoic acid and 9.21 mL (6.68 g, 66.7 mmol, 1.33 equiv) of triethylamine (freshly distilled from CaH2 under N2) in 100 mL of dry THF, cooled to -78 °C under No. was added 7.08 ml. (6.93 o 57.5 mmol 1.15 equiv) of nivelevi chloride. The mixture was warmed to 0.97 for 60 min and then recooled to -78 °C. A solution of 8.86 g (50.0 mmol, 1.00 equiv) of (45)-(phenylmethyl)-2-oxazolidone (2) (XpH) and 15 mg of triphenylmethane (indicator) in 90 mL of dry THF, stirred at -30 °C to -45 °C under N2, was treated dropwise with n-butyllithium (2.48 M in hexane) until an orange color persisted (20.2 mL, 1.00 equiv required). The resulting solution was cooled to -78 °C and then added, via rapid cannulation, to the above stirred mixture containing the mixed anhydride. The residual metalated oxazolidone was rinsed in with two 10-mL portions of dry THF and the resulting mixture was stirred at -78 °C for 30 min. After warming to 0 °C, the mixture was partitioned between CH2Cl2 and pH 7 phosphate buffer. The CH2Cl2 phase washed with 5% aqueous NaHCO3 followed by half-saturated aqueous NaCl, dried (MgSO4), and evaporated in vacuo. The residual oil was chromatographed on two size D Michel-Miller columns (660 g of silica gel) eluting with 3: 1 hexane-EtOAc. Mixed fractions (~2 g) were rechromatographed as described above to give a baseline separation of 3b as a colorless, viscous oil (11.5 g) that was homogeneous on TLC: Rf 0.27 (silica, 3: 1 hexane-EtOAc). Kugelrohr distillation (150 °C/8 millitorr) yielded 11.4 g (87%) of pure material: IR (neat) 3065, 3025, 2975, 2920, 1782, 1702, 1387, 1352, 1210 cm⁻¹; ¹H NMR (250 MHz, CDCl₃) 8 7.37-7.19 (m, 5H, C₆H₅), 5.884 (ddx, $J_{cis} = 10.3 \text{ Hz}$, $J_{trans} = 17.0 \text{ Hz}$, $J_{allyl} = 6.5 \text{ Hz}$, 1H, $H_2C = CHCH_2$), 5.108 (dm, $J_{trans} = 17.0 \text{ Hz}$, 1H, $H_cC = CHCH_2$), 5.032 (dm, J_{Cis} = 10.3 Hz, 1H, H_CC=CHCH₂), 4.716-4.622 (sym m, 1H, CH₂CH_N), 4.227-4.123 (m, 2H, CH₂O), 3.288 (dd, J = 3.3, 13.4 Hz, 1H, CHHC₆H₅), 3.169-2.935 (AB quartet of triplets, $v_A = 3.091$, $v_B = 3.011$, $J_{AB} = 17.3$ Hz, $J_t =$ 7.4 Hz, 2H, CHHC=O), 2.762 (dd, J = 9.6, 13.4 Hz, 1H, CHHC6H5), 2.502-2.411 (m, 2H, H2C=CHCH2); ¹³C NMR (62.9 MHz, CDCl₃) & 172.44, 153.30, 136.69, 135.35, 129.31, 128.85, 127.24, 115.51, 66.19, 55.09, 38.02, 34.80, 28.21; $[\alpha]_D^{25}$ +62.0° (c = 1.13, CHCl₃). Anal. Calcd for C₁₅H₁₇NO₃: C, 69.48; H, 6.61. Found: C, 69.57; H, 6.59.

(4S)-3-(1-oxo-3-phenylpropyl)-4-(phenylmethyl)-2-oxazolidinone (3c, R = CH₂C₆H₅). A solution of 22.55 g (127.3 mmol, 1.00 equiv) of (4S)-(phenylmethyl)-2-oxazolidone (2) (XpH) and 50 mg of triphenylmethane (indicator) in 250 mL of dry THF, stirred at -78 °C under N₂, was treated dropwise with n-butyllithium (1.55 M in hexane) until an orange color persisted (82 mL, 1.00 equiv required); the solution being kept at -65 to -78 °C during the addition. After recooling to -78 °, the solution was treated with 21.0 mL (23.8, 141 mmol, 1.10 equiv) of dihydrocinnamoyl chloride over a 2-min period. The resulting mixture was warmed to 0 °C, treated with 100 mL of saturated aqueous NaHCO₃, and stirred at 20 °C for 30 min. The mixture was extracted with three portions of CH₂Cl₂ extracts were combined and successively washed with 5% aqueous Na₂CO₃ and saturated aqueous NaCl, dried (MgSO₄), and evaporated in vacuo to afford an off-white solid. This material was recrystallized from 450 mL of 2: 1 hexane-EtOAc to yield 36.4 g (92%) of glistening white needles: TLC R₁ 0.27 (silica, 3: 1 hexane-EtOAc); mp 109-109.5 °C; IR (CHCl₃)

1780, 1695 cm $^{-1}$; 1 H NMR (300 MHz, CDCl3) & 7.35 (m, 10H, aromatics), 4.700-4.622 (sym m, 1H, CH₂CHN), 4.209-4.126 (m, 2H, CH₂O), 3.382-3.187 (m, 3H, CH₂C=O and 4-CHHC₆H₅), 3.102-2.952 (m, 2H, CH₂CH₂C=O), 2.751 (dd, J = 9.5, 13.4 Hz, 1H, 4-CHHC₆H₅); 13 C NMR (75.5 MHz, CDCl3) & 172.41, 153.42, 140.73, 135.57, 129.57, 129.05, 128.77, 128.60, 127.43, 126.40, 66.19, \$5.25, 38.05, 37.27, 30.60; $[\alpha]_{D}^{25}$ +78.6° (c = 1.00, CH₂Cl₂). Anal. Calcd for C₁9H₁9NO₃: C, 73.75; H, 6.20. Found: C, 73.70; H, 6.21.

(4S)-3-(phenylacetyl)-4-(phenylmethyl)-2-exazolidinone (3d, R = C6H5). A solution of 12.4 g (70.0 mmol, 1.00 equiv) of (4S)-(phenylmethyl)-2-oxazolidone (2) (XpH) and 25 mg of triphenylmethane (indicator) in 130 mL of dry THF, stirred at -78 °C under N2, was treated dropwise with n-butyllithium (2.55 M in hexane) until an orange color persisted (27.5 mL, 1.00 equiv required); the solution being kept at -65 to -78 °C during the addition. After recoeling to -78 °C, the solution was treated with 10.0 mL (11.5 g, 74.1 mmol, 1.06 equiv) of phenylacetyl chloride (98%) over a 2-min period. The resulting mixture was warmed to 25 °C, treated wish 100 mL of saturated aqueous NaHCO3, and stirred at 25 °C for 30 min. The mixture was extracted with three portions of CH2Cl2. The CH2Cl2 extracts were combined, washed with 5% aqueous Na₂CO₃, washed with saturated aqueous NaCl, dried (MgSO₄), and evaporated in vacuo. The residual yellow oil was triturated with three portions of Et2O to afford, after thorough air drying, 8.73 g (42%) of 3d as a creamcolored solid found by TLC to contain a trace of XpH. The triturate liquors were combined and evaporated in wacuo. The residual yellow oil (11.1 g) was chromatographed on two size D Michel-Miller columns (660 g of silica gel) eluting with 3: 1 hexane-EtOAc to yield an additional 7.05 (34%) of 3d, found by TLC (hexane-EtOAc (65: 35)) to contain a trace faster moving impurity. The two lots were combined and recrystallized from Et2O-hexane to afford 11.6 g of 3d (white needles) that was homogeneous on TLC: Re 0.54 (silica, 1: 1 hexane-EtOAc); mp 71.5-72.5 °C. IR (CHCl3) 3030, 1783, 1700, 1498, 1454, 1385, 1362, 1108 cm⁻¹; ³H NMR (300 MHz, CDCl₃) δ 7.39-7.12 (m, 10H, aromatics), 4.719-4.641 (sym m, 1H, CH₂CHN), 4.388-4.294 (AB quartet, $v_A = 4.343$, $v_B = 4.277$, $J_{AB} = 15.7$ Hz, 2H, CH₂C=O), 4.21-4.15 (m, 2H, CH₂O), 3.275 (dd, J = 3.2, 13.4 Hz, 1H, 4-CHHC₆H₅), 2.756 (dd, J = 9.5 13.4 Hz, 1H, 4-CHHC₆H₅); ¹³C NMR (75.5 MHz, CDCl₃) δ 170.93, 153.19, 135.01, 133.47, 129.61, 129.24, 128.71, 128.37, 127.09, 127.03, 65.90, 55.02, 41.33, 37.48; $[\alpha]_0^{22}$ +72.4° (c = 1.02, CHCl3). Anal. Calcd for C₁₈H₁₇NO₃: C, 73.20; H, 5.80. Found: C, 73.02; H, 5.72.

(4S)-3-(3-methyl-1-oxobutyl)-4-(phenylmethyl)-2-oxazolidinone (3e, R = CH(CH₃)₂). A solution of 8.86 g (50.0 mmol, 1.00 equiv) of (45)-(phenylmethyi)-2-oxazolidone (2) (XpH) and 15 mg of triphenylmethane (indicator) in 90 mL of dry THF, stirred at -50 °C under N2, was treated dropwise with n-butyllithium (~2.5 M in hexane) until an orange color persisted (19.9 mL, 1.00 equiv required); the solution warming to -30 °C during the addition. After recooling to -78 °C, the solution was treated with 6.71 mL (6.64 g, 55.0 mmol, 1.10 equiv) of redistilled isovaleryl chloride in one portion. The resulting solution was stirred at -78 °C for 15 min, warmed to 0 °C, and treated with 100 mL of saturated aqueous NaHCO3. The mixture was stirred vigorously at 20 °C for 45 min, and was then partitioned between CH2Cl2 and H2O containing 15 mL of 10% aqueous Na2CO3. The aqueous phase was extracted with three additional portions of CH2Cl2. The CH2Cl2 extracts were combined, washed with saturated aqueous NaCl, dried (MgSO₄), and evaporated in vacuo to give 13.5 g of a pale-yellow oil. This material was crystallized from Et2O-hexane to yield, in three crops, 12.0 g (92%) of 3e as fine white needles: TLC Rf 0.40 (silica, 7: 3 hexane-EtOAc); mp 50.0-51.0 °C; IR (CHCl₃) 3030, 2962, 2870, 1782, 1698, 1390, 1385, 1352 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.37-7.21 (m, 5H, $C_{6}H_{5}$), 4.725-4.646 (sym m, 1H, $CH_{2}C_{H}N$), 4.225-4.133 (m, 2H, $CH_{2}O$), 3.316 (dd, J = 3.3, 13.3 Hz, 1H, $C_{H}HC_{6}H_{5}$), 2.936-2.738 (8-line AB portion of ABX system, $v_A = 2.891$, $v_B = 2.784$, $J_{AB} = 16.2$ Hz, $J_{AX} = 6.7$ Hz, $J_{BX} = 6.9$ Hz, $CH_2C=O$) and 2.752 (dd, J=9.7 13.3 Hz, $CH_2H_2GH_3$) (3H total), 2.225 (sym 9-line m, J=6.7 Hz, 1H, $CH_3(CH_3)_2$), 1.025 (d, J = 6.7 Hz) and 1.009 (d, J = 6.7 Hz) (6H total); ¹³C NMR (75.5 MHz, CDCl₃) & 172.52, 153.30, 135.34, 129.29, 128.81, 127.20, 66.01, 55.02, 43.87, 37.93, 24.96, 22.42, 22.31; $[\alpha]_D^{25}$ +55.8° (c = 1.01, CHCl₃). Anal. Calcd for C₁₅H₁₉NO₃: C, 68.94; H, 7.33. Found: C, 68.87; H, 7.30.

(4S)-3-(3,3-dimethyl-1-oxobutyl)-4-(phenylmethyl)-2-oxazolidimone (3f, R = C(CH₃)₃). A solution of 6.44 g (36.3 mmol, 1.00 equiv) of (4S)-(phenylmethyl)-2-oxazolidone (2) (XpH) in 70 mL of dry THF, stirred at 78 °C under N₂, was treated dropwise with 23.1mL (1.0 equiv) of n-butyllithium (1.56 M in became) over a 5-min period. The solution was stirred at -78 °C for 15 min, and then treated dropwise with 5.50 mL (5.33 g, 39.6 mmol, 1.09 equiv) of t-butylacetyl chloride. After stirring at -78 °C for 20 min, the solution was warmed to 25 °C over a 1-h period and treated with 10 mL of saturated aqueous NH₄Cl. The THF was evaporated in vacuo. The aqueous residue was extracted with three 80-mL portions of Et₂O. The Et₂O extracts were combined, washed with 1 N NaOH, washed with saturated aqueous NaCl, dried (Na₂SO₄), and evaporated in vacuo. The residual white solid was recrystallized from Et₂O to yield, in two crops, 9.23 g (92%) of 3f as fine white needles: mp 112.5-113.5 °C; IR (CHCl₃) 3020, 2960, 2870, 1780, 1690, 1385, 1362, 1353 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.37-7.22 (m, 5H, C₆H₅), 4.738-4.695 (sym m, 1H, CH₂CHN), 4.192-4.122 (m, 2H, CH₂O), 3.350 (dd, J = 3.3, 13.3 Hz, 1H, CHHC₆H₅), 3.023-2.836 (AB quartet, ν_A = 2.994, ν_B = 2.866, J_{AB} = 14.9 Hz, 2H, CH₂C=O), 2.713 (dd, J = 10.0, 13.3 Hz, 1H, CHHC₆H₅), 1.096 (s, 9H, C(CH₃)₃); ¹³C NMR (75.5 MHz, CDCl₃) δ 171.79, 153.39, 135.43, 129.32, 128.84, 127.20, 65.73, 55.25, 46.08, 38.01, 31.32, 29.53; α ₁D²2+42.5° (c = 1.02, CHCl₃). Anal. Calcd for C₁6H₂1NO₃: C, 69.79; H, 7.69. Found: C, 69.96; H, 7.75.

(4S)-3-(1,5-diexo-5-methoxypentyl)-4-(phenylmethyl)-2-exazolidinone (3g, R = CH₂CH₂CO₂. Me). A solution of 8.86 g (50.0 mmoi, 1.00 equiv) of (4\$)-(phenylmethyl)-2-oxazolidone (2) (XpH) and 15 mg of triphenylmethane (indicator) in 90 mL of dry THF, stirred at -50 °C under N2, was treated dropwise with n-butyllithium (~2.5 M in hexane) until an orange color persisted (19.9 mL, 1.00 equiv required); the solution warming to -30 °C during the addition. After recooling to -78 °C, the solution was treated with 7.60 mL (9.05, 55.0 mmol, 1.10 equiv) of methyl 4-(chloroformyl)butyrate in one portion. The resulting solution was stirred at -78 °C for 15 min, warmed to 0 °C, and treated with 100 mL of saturated aqueous NaHCO₃. The mixture was stirred vigorously at 20 °C for 45 min, and was then partitioned between CH2Cl2 and H2O containing 15 mL of 10% aqueous Na2CO3. The aqueous phase was extracted with three additional portions of CH2Cl2. The CH2Cl2 extracts were combined, washed with saturated aqueous NaCl, dried (MgSO₄), and evaporated in vacuo to give 15.9 g of a pale-yellow oil which solidified on standing. One recrystallization from hexane-EtOAc yielded 10.8 g (71%) of white solid (first crop) that was homogeneous on TLC: Rf 0.23 (silica, CH2Cl2-bexane-CH3CN (70: 30: 5)); mp 72.5-74.0 °C; IR (CHCl3) 3030, 3020, 2955, 1782, 1732, 1700, 1385, 1355 cm⁻¹; ¹H NMR (250 MHz, CDCl₃) 8 7.38-7.19 (m, 5H, C₆H₅), 4.718-4.624 (sym m, 1H, CH₂CHN), 4.250-4.142 (m, 2H, CH₂O), 3.690 (s, 3H, CO₂CH₃), 3.300 (dd, J = 3.3, 13.3 Hz, 1H, CHHC₆H₅), 3.107-2.895 (sym m, 2H, CH_2CH_2CON), 2.771 (dd, J = 9.6, 13.3 Hz, 1H, $CH_1H_2GH_3$), 2.443 (t, J = 7.3 Hz, 2H, $CH_2CO_2CH_3$, 2.094-1.640 (m, 2H, CH₂CH₂CH₂); ¹³C NMR (62.9 MHz, CDCl₃) 8 173.07, 172.25, 153.22, 135.18, 129.21, 128.73, 127.13, 66.13, 54.92, 51.29, 37.79, 34.51, 32.91, 19.38; $\{\alpha\}_{D}^{22} + 51.0^{\circ} (c = 1.02, CHCl_3)$. Anal. Calcd for $C_{16}H_{19}NO_{5}$: C, 62.94; H, 6.27. Found: C, 62.88; H, 6.36.

(4S)-3-(3-methyl-1-oxo-2-butenyl)-4-(phenylmethyl)-2-oxazolidinone (3h). A solution of 8.93 g (50.4 mmol, 1.00 equiv) of (4S)-(phenylmethyl)-2-oxazolidone (2) (XpH) in 100 mL of dry THF, stirred at -78 °C under N₂, was treated dropwise with 20.9 mL (53.9 mmol, n-butyllithium (2.58 M in hexane) over a 5-min period. The solution was stirred at -78 °C for an additional 5 min, and then treated with 6.00 mL (6.39 g, 53.9 mmol, 1.07 equiv) of 3,3-dimethylacryloyl chloride (97%) in one portion. The resulting solution was stirred at -78 °C for 5 min, warmed to 25 °C, and treated with 5 mL of saturated aqueous NH₄Cl. The THF was evaporated in vacuo, and the residue was partitioned between 300 mL of 1:1 CH₂Cl₂-hexane and 100 mL of saturated aqueous NaHCO₃. The organic phase was washed with saturated aqueous NaCl, dried (Na₂SO₄), and evaporated in vacuo to yield 13.1 g of white solid. This material was recrystallized from hexane-EtOAc to yield, in two crops, 11.5 g (88%) of 3h as fine white needles: TLC R_f 0.60 (silica, 1: 1 hexane-EtOAc); mp 79.5-80.5 °C; IR (CHCl₃) 3020, 2920, 1772, 1677, 1630, 1385, 1360, 1350, 1257, 1184 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.37-7.22 (m, 5H, C₆H₅), 6.966-6.950 (m, 1H, C=CHC=O), 4.762-4.684 (sym m, 1H, CH₂CH₁N), 4.213-4.118 (m, 2H, CH₂O), 3.347 (dd, 1 = 3.3, 13.3 Hz, 1H, CHHC₄H₅), 2.775 (dd, 1 = 9.7, 13.3 Hz, 1H, CH₂CH₃O), 2.230 (g, z = 1.2 Hz, 3H, CH₂O), 2.347 (dd, 1 = 3.3, 13.3 Hz, 1H, CHHC₄H₅), 2.775 (dd, 1 = 9.7, 13.3 Hz, 1H, CH₂CH₃O), 2.230 (g, z = 1.2 Hz, 3H, CH₂O), 2.28.84, 127.17, 115.79, 65.84, 55.14, 38.01, 28.00, 21.31; [α]_D²² +60.4° (c = 1.02, CHCl₃). Anal. Calcd for C₁5H₁7NO₃: C, 69.48; H, 6.61. Found: C, 69.56; H, 6.63.

General Procedure for the Formation of the Carboxinside (Z) Lithium Enolates and Their Subsequent Reaction with Di-f-butyl Azodicarboxylate. To a freshly prepared solution of 1.05 mmol of lithium disopropylamide in 3 mL of THF, stirred at -78 °C under N₂, was added via cannula a precooled (-78 °C) solution of 1.00 mmol of the imide, 3, in 3 mL of THF. Residual 3 was rinsed in with two 1 mL portions of THF and stirring continued at -78 °C for 30 min. A precooled (-78 °C) solution of 265 mg (1.15 mmol) of DBAD in 6 mL of dry CH₂Cl₂ was added via cannula to the above enolate solution and after an additional 30-180 sec the reaction was quenched with 2.6 mmol of glacial acetic acid. The mixture was partitioned between CH₂Cl₂ and pH 7 phosphate buffer. The aqueous phase was washed with three portions of CH₂Cl₂. The CH₂Cl₂ phases were combined, washed with saturated aqueous NaHCO₃, dried (MgSO₄), and evaporated in vacuo. Diastereometic ratios of the resulting unpurified product were obtained by HPLC analysis on a Waters, 8mm x 10 cm 5 micron silica gel Radial-Pak^R column (UV detection at 235 nm) eluting with either CH₂Cl₂-hexane-CH₃CN (70:30:5) (Solvent A) or CH₂Cl₂-hexane-CH₃CN (70:30:7.5) (Solvent B) (vide infra) at 2 mL/min. The unpurified product was purified by medium pressure chromatography (MPLC) on Michel-Miller columns (Ace Glass) packed with silica gel (Merck spectophotometer 230-400 mesh) with typical column loadings of 0.3 to 1 g of material per 100 g of adsorbent (see below for solvent):

(3(2S),4S)-3-(2-(N,N'-bis-(t-butoxycarbomyl)hydrazino)-1-oxopropyl)-4-(phenylmethyl)-2-oxazolidinome (4a, R = CH3) (Table 1, Entry A). As described above, 233 mg (1.00 mmol) of 3a (R = CH3)³⁷ afforded 424 mg (92%) of 4a (R = CH3) as a colorless glass foam after purification by MPLC (50 g of silica gel; CH2Cl2-hexane-CH3CN (70:30:7)). Recovery of 3a : 13.2 mg (6%). Diastereomer analysis of the unpurified product (vide supra, solvent A) gave a 2(S) (t_T = 4.89 min): 2(R) (minor diastereomer, t_T = 6.24 min) ratio of 98:2. The purified product gave the same diastereomer ratio: IR (CHCl3) 3400, 1788, 1747, 1705 cm⁻¹; ¹H NMR (300 MHz, CDCl3, 329 °K) δ 7.35-7.19 (m, 5H, aromatics), 6.52 (br s, 1H, NH), 5.74 (br s, 1H, CHC=O), 4.63-4.56 (sym 8 line m, 1H, 4-H), 4.22-4.13 (m, 2H, OCH2), 3.339 (dd, J_{ViC} = 2.8 Hz, J_{gem} = 13.4 Hz, 1H, CHHPh), 2.754 (dd, J_{ViC} = 9.9 Hz, J_{gem} = 13.4 Hz, 1H, CHHPh), 1.482 (s, OC(CH3)3), 1.466 (s, OC(CH3)3) and 1.453 (s, CHCH3)(21H total); $[\alpha]_D^{26}$ +37.7° (c = 1.05, CHCl3). Anal. Calcd for C23H33N3O7: C, 59.60; H, 7.18. Found: C, 59.54; H, 7.13.

Preparation of the Minor Diastersomer (2(R)-4a, R = CH3) Vin Epimerization. A solution of 0.400 g (0.863 mmol) of 4a (R = CH3) and 0.23 mL (0.21g; 1.83 mmol; 2.1 equiv) of 1,1,3,3-ectamethylguanidine in 9 mL of dry THF was refluxed under N2 for 4.5 h. The solution as partitioned between aqueous pH 7 buffer and CH2Cl2. The aqueous phasewas washed with CH2Cl2 (x 2), and the organic phases were combined, dried (MgSO4), and evaporated in vacuo. The unpurified product was purified by flash chromatography (45 g of silica gel, hexane-BrOAc (1:1)) to give 330 mg (82% recovery) of a coloriest glass, assayed by HPLC (vide supra) to have a 2(5):2(R) ratio of 56:44. HNMR (300 MHz, CDCl3, 329 °K) integration of the low field diasterectopic benzylic protons afforded a 2(5):2(R) ratio of 54:46. Distinctive HNMR signals attributed to the minor diastereomer (2(R)-4a) include the following: 8 4.71-4.62 (m, 1H, 4-H), 4.28-4.23 (low field half of AB m, OCHH), 3.223 dd, Jvic = 3.4 Hz, Jgem = 13.4 Hz, PhCHH), 2.788 (dd, Jvic = 9.2 Hz, Jgem = 13.5 Hz, PhCHH).

(3(2S),4S)-3-(2-(N,N'-bis-(t-butoxycarbonyl))) hydraxino)-1-oxo-4-pentenyl)-4-(phenylmethyl)-2-oxazolidinone (4b, R = CH₂CH=CH₂) (Table 1, Entry B). Pollowing an exact scale up of the general procedure, 2.59 g (10.0 mmol) of 3b (R = CH₂CH=CH₂) gave 4.59 g (94%) of 4b (R = CH₂CH=CH₂) as a colorless glass foam after purification by MPLC (650 g of silica gel; Solvent A). HPLC analysis (vide supra, Solvent A) of the unpurified product gave a 2(S) (t_f = 5.79 min): 2(R) (t_f = 8.23 min) ratio of 98:2. HPLC analysis of the purified product gave a 2(S):2(R) ratio of >300:1 : IR (CH₂Cl₂) 3385, 1785, 1750, 1710 cm⁻¹; ¹H NMR (250 MHz, CDCl₃, 331 °K) δ 7.36-7.19 (m, 5H, aromatic), 6.52 (br s, 1H, NH), 5.932 (ddt, t_{cis} = 10.2 Hz, t_{trans} = 17.1 Hz, t_{trans} = 17.1

Later fractions gave 94 mg (1.9%) of the minor diastereomer 2(R)-4c (R = CH₂CH=CH₂) assayed by HPLC to have a 2(R):2(S) ratio of 99:1 : IR (CH₂Cl₂) 3385, 1785, 1755, 1710 cm⁻¹; ¹H NMR (250 MHz, CDCl₃, 331 °K) & 7.35-7.16 (m, 5H, aromatic), 6.520 (br s, 1H, NH), 5.962 (ddx, J_{Cis} = 10.1 Hz, J_{trans} = 17.1 Hz, J_{allyl} = 6.94 Hz, 1H, C=CHCH₂), 5.828 (apparent t, 1H, J = 6.7 Hz, CHC=O), 5.198 (dm, J = 17.1 Hz, 1H, H_{c} C=CHCH₂), 5.086 (dm, J = 10.1 Hz, 1H, H_{c} C=CHCH₂), 4.68-4.58 (sym m, 1H, 4-H), 4.23 (apparent t, J = 8.4 Hz, 1H, OCHH), 4.14 (dd, J = 3.3, 9.0 Hz, 1H, OCHH), 3.273 (dd, J_{vic} = 3.4 Hz, J_{gem} = 13.5 Hz, 1H, CHHC₆H₅), 2.76 -2.55 (m, 3H, CHHC₆H₅ and C=CHCH₂), 1.474 (s) and 1.451 (s) (18H, OC(CH₃)₃).

(3(2S),4S)-3-(2-(N,N'-bis-(t-butoxycarbonyl)hydraxino)-3-phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl)-4-(phenyl-1-oxopropyl-1-oxopropyl-1-oxopropyl)-4-(phenyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxopropyl-1-oxomethyl)-2-oxazolidinone (4c, R = CH2Ph) (Table 1, Entry C). Following a modification of the general procedure, a solution of 6.19 g (20.0 mmol) of 3c (R = CH₂C₆H₅) in 60 mL of THF was added via cannula over a 50 min period to a stirred, precooled (-78 °C) solution of 21.0 mmol of LDA in 20 mL of THF and 10 mL of hexane. Residual 3c was rinsed in with two 5 mL portions of THF and enolization continued at -78 °C for 30 min. The above precooled (-78 °C) solution of lithium enolate was added via cannula to a stirred, precooled (-78 °C) solution of 5.30 g (23.0 mmol) of DBAD in 120 mL of CH2Cl2 over a 9 min period. Residual enolate was rinsed in with two 10 mL portions of THF, and after an additional 60 sec the reaction was quenched with 3.0 mL (52 mmol) of glacial acetic acid. Following the standard workup (vide supra) and purification by MPLC (900 g of silica gel, Solvent A) 9.83 g (91%) of 2 (R = CH2C6H5) was obtained as a colorless glass foam. HPLC analysis (vide supra, Solvent A) of the unpurified product gave a 2(S) ($t_{\rm f} = 7.07$ min): 2(R) (t_r = 10.76) ratio of 97:3. HPLC analysis of the purified product gave a 2(S):2(R) ratio >400:1: IR (CH₂Cl₂) 3390, 1785, 1750, 1712 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 330 °K) & 7.330-7.136 (m, 10H, aromatics), 6.50 (br s, 1H, NH), 6.15 (br s, 1H, CHC=O), 4.45-4.37 (sym m, 1H, 4-H), 4.008 (dd, $J_{vic} = 2.8$, $J_{gem} = 8.9$ Hz, 1H, OCHH), 3.832 (apparent t, J = 8.3 Hz, 1H, OCHH), 3.303 (dd, $J_{vic} = 3.0$ Hz, $J_{gem} = 13.6$ Hz, 1H, NCHCHHPh) 3.25-3.15 (AB m, 2H, PhCH2CHC=O), 2.685 (dd, J_{vic} = 9.9 Hz, J_{gem} = 13.5 Hz, 1H, NCHCHHPh), 1.452 (s) and 1.444 (s) (18H, OC(CH₃)₃); $[\alpha]_D^{26}$ +84.4° (c = 1.01, CHCl3). Anal. Calcd for C₂₉H₃₇N₃O₇: C, 64.55; H, 6.91. Found: C, 64.48; H, 6.90.

After several overlapping fractions (317 mg; 2.9%) there was obtained 254 mg (2.4%) of the minor diastereomer, 2(R)-4c (R = CH₂C6H₅) which was found by HPLC (vide supra) to have a 2(R):2(S) ratio of 98:2: IR (CH₂Cl₂) 3385, 1788, 1753, 1713 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 329 °K) δ 7.34-7.08 (m, 10H, aromatics), 6.386 (br s, 1H, NH), 6.165 (br t, J = 6.8 Hz, 1H, CHC=O), 4.61-4.55 (m, 1H, 4-H), 4.205 (apparent t, J = 8.4 Hz, 1H, OCHH), 4.070 (dd, J_{vic} = 3.0 Hz, J_{gem} = 8.9 Hz, 1H, OCHH), 3.28-3.13 (m, 3H, PhCH₂CHC=O and NCHCHHPh), 2.590 (dd, J_{vic} = 9.5 Hz, J_{gem} = 13.5 Hz), 1.409 (s, 18H, OC(CH₃)₃).

 $(3(2S),4S)-3-(2-(N_rN'-bis-(t-butoxycarbonyl)hydrazino)phenylacetyl)-4-(phonylmethyl)-2-oxazolidinone (4d, R = Ph) (Table 1, Entry D). Following the general procedure, 295 mg (1.00 mmol) of 3d (R = Ph) afforded 503 mg (96%) of 4d (R = C₆H₅) as a colorless glass foam after purification by MPLC (170 g of silica gel, Solvent B). Diastereomer analysis of the unpurified product by HPLC (Solvent B, vide supra) gave a 2(S) (<math>t_r = 5.66$ min): 2(R) ($t_r = 8.71$ min) ratio of 97:3. HPLC analysis of the purified product gave a 2(S):2(R) ratio of >400:1: IR

(CH₂Cl₂) 3400, 1788, 1750, 1700 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 329 °K) 7.38-7.22 (m, 10H, aromatics), 7.06 (s, 1H), 6.49 (br s, 1H), 4.65-4.57 (sym m, 1H, 4-H), 4.12-4.01 (m, 2H, OCH₂), 3.379 (br d, 1H, PhCHH), 2.848 (dd, J_{vlc} = 9.7 Hz, J_{gem} = 13.3 Hz, 1H, PhCHH) 1.500 (s, 9H, OC(CH₃)₃), 1.191 (br s, 9H, OC(CH₃)₃); [α]_D²⁶ +216° (c = 1.04, CHCl₃). Anal. Calcd for C₂₈H₃SN₃O₇: C, 63.98; H, 6.71. Found: C, 64.07; H, 6.62.

Later fractions afforded 13 mg (2.4%) of the minor diastereomer 2(R)-4d ($R = C_6H_5$), shown by HPLC analysis to have a 2(R): 2(S) ratio of 97:3 : IR (CH₂Cl₂) 3405, 1785, 1750, 1700 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 330 °K) δ 7.43-7.22 (m, 8H, aromatics), 7.09-7.05 (m, 3H, 2 aromatics and NH or CHC=O), 6.50 (br s, 1H), 4.79-4.71 (sym m, 1H, 4-H), 4.190 (apparent t, J = 8.6 Hz, 1H, OCHH), 4.052 (dd, $J_{\rm vic} = 3.5$ Hz, $J_{\rm gem} = 9.1$ Hz, 1H, OCHH), 3.320 (dd, $J_{\rm vic} = 3.2$ Hz, $J_{\rm gem} = 13.5$ Hz, 1H, PhCHH), 1.481 (s, 9H, OC(CH₃)₃), 1.194 (br s, 9H, OC(CH₃)₃).

(3(2S),4S)-3-(2-(N,N'-bis-(i-butoxycarbonyl))) hydrazino)-3-methyl-1-oxebutyl)-4-(phenyl-methyl)-2-oxazolidinone (4e, R = i-Pr) (Table 1, Entry E). Following an exact scale up of the general procedure, 2.61 g (10.0 mmol) of 3e (R = i-Pr) gave 4.65 g (95%) of 4e (R = i-Pr) as a colorless glass foam after purification by MPLC (650 g of silica gel; Solvent A). HPLC analysis (vide supra, Solvent A) of the unpurified product gave a 2(S) ($t_T = 5.77$ min): 2(R) ($t_T = 8.08$ min) ratio of 98:2. HPLC analysis of the purified product gave a 2(S): 2(R) ratio >400: 1: IR (CH₂Cl₂) 3390, 1785, 1750, 1708 cm⁻¹; ¹H NMR (250 MHz; CDCl₃; 331 °K) δ 7.35-7.19 (m, 5H, aromatics), 6.48 (br s, 1H, NH), 5.79 (br d, J = 8.2 Hz, 1H, CHC=0, 4.685-4.588 (m, 1H, 4-H), 4.192-4.115 (m, 2H, OCH₂), 3.363 (br dd, $J_{vic} = 3$ Hz, $J_{gem} = 13.5$ Hz, 1H, CHHC₆H₅), 2.718 (dd, $J_{vic} = 9.9$ Hz, $J_{gem} = 13.5$ Hz, 1H, CHHC₆H₅), 2.412-2.267 (m, 1H, CH(CH₃)₂), 1.478 (s) and 1.473 (s) (18H, OC(CH₃)₃), 1.099 (d, J = 6.7 Hz, 3H, CHCH₃), 1.000 (d, J = 6.9 Hz, 3H, CHCH₃); $\{\alpha\}_D^{26} + 27.4^{\circ}$ (c = 1.02, CHCl₃). Anal. Calcd for C₂₅H₃₇N₃O₇: C, 61.08; H, 7.59. Found: C, 60.90; H, 7.51.

Preparation of the minor diastereomer (2(R)-4e, R=i-Pr) via Epimerization. The protocol for carrying out the epimerization was the same as that described above for 4e, (R=Me). The reaction time was 19 h.

(3(2S),4S)-3-(2-(N,N'-bis-(t-butoxycarbomyl)hydrazino)-3,3-dimethyl-1-oxobutyl)-4-(phenyl-methyl)-2-oxazolidinone (4f, R = t-Bu) (Table 1, Entry F). Following a slightly modified (40 min enolization time) scale up of the general procedure, 2.75 g (10.0 mmol) of 3f (R = t-Bu) gave 4.83 g (96%) of 4f (R = t-Bu) as a colorless glass foam after purification by MPLC (vide supra, Solvent A). HPLC analysis (vide supra, Solvent A) of the unpurified product gave a 2(S) (t_f = 5.33 min): 2(R) (t_f = 7.73 min) ratio of 99.7:0.3. HPLC analysis of the purified product gave a 2(S):2(R) ratio >500:1 : IR (CH₂Cl₂) 3395, 1785, 1748, 1710 cm⁻¹; ¹H NMR (250 MHz, CDCl₃, 331 °K) δ 7.34-7.20 (m, 5H, aromatics), very broad singlets at 6.45 and 6.07 (2H, NH and CHC=O), 4.71-4.61 (m, 1H, 4-H), 4.16-4.10 (m, 2H, OCH₂), 3.45-3.35 (unresolved m, 1H, CHHC₆H₅), 2.696 (dd, J_{Vic} = 10.2 Hz, J_{gem} = 13.4 Hz, 1H, CHHC₆H₅), 1.491 (s) and 1.452 (s) (18H, OC(CH₂)₂), 1.116 (s, 9H, CC(CH₂)₂); I_{CH} [c) +13.4° (c) = 1.01, CHCl₂). Anal.

(3(2S),4S)-3-(2-(N,N'-bis-(t-butoxycarbonyl)hydrazino)-5-methoxy-1,5-dioxopentyl)-4-(phenylmethyl)-2-oxazolidinome (4g, R = CH2CH2CO2Me) A solution of 305 mg (1.00 mmol, 1.00 equiv) of 3g in 3 ml of dry THF, stirred at -78 °C under N2, was treated via rapid cannulation with a precooled (-78 °C) solution of 1.01 mmol of LDA (prepared as described above from 0.64 mL (1.01 mmol, 1.01 equiv) of n-butyllithium (1.58 M in hexane) and 0.15 mL (1.07 mmol) of diisopropyl amine in 3 mL of THF). Residual LDA was rinsed in with one 2-mL portion of THF, and the resulting solution was stirred at -78 °C for 5 min. To the above was added, via rapid cannulation, a precooled (-78 °C) solution of 265 mg (1.15 mmol, 1.15 equiv) of DBAD in 6 mL of CH2Cl2. After 40 sec the reaction was quenched with 0.15 mL (157 mg, 2.6 mmol) of glacial HOAc. The unpurified product obtained following the standard workup was purified by MPLC (106 g of silica gel; 2L linear gradient from Solvent A to CH2Cl2-hexane-CH3CN (70: 30: 15)) to afford, in order of elution, 47.6 mg (16% recovery) of 3g followed by 267 mg (51% yield) of 4g as a colorless glass foam: IR (CHCl3) 3385, 3060, 2973, 1785, 1750-1700 (br. max at 1735), 1480, 1393, 1369, 1353, 1235, 1197, 1173, 1152, 1110 cm⁻¹; ¹H NMR (250 MHz, CDCl₃, 331 °K) & 7.36-7.17 (m, 5H, C₆H₅), 6.59 (br s, 1H, NH), 5.70 (br d, J = 8.6 Hz, 1H, CHC=O), 4.63-4.53 (sym m, 1H, CH2CHN), 4.24-4.13 (m, 2H, CH2O), 3.638 (s, 3H, CO2CH3), 3.30 (br d, J ~ 13.6 Hz, 1H, CHHC6H5), 2.88-2.73 (m, 2H, CHHC6H5 and CHHCO2CH3), 2.54 (ddd, 1H, J = 6.3, 9.7, 16.5 Hz, 1H, CHHCO₂CH₃), 2.29-1.97 (m, 2H, CH₂CH₂CHN), 1.475 (s, 18H, OC(CH₃)₃); $\{\alpha\}_D^{26}$ +46.9° (c = 1.02, CHCl3). Anal. Calcd for C26H37N3O9: C, 58.31; H, 6.96. Found: C, 58.36; H, 7.09. This material was found to be homogeneous on HPLC: tr 13.85 min (Solvent A), tr 7.59 min (Solvent B). H NMR analysis indicates a minimum diastereomeric purity of >95: 5 (vide infra).

Preparation of the minor diastereomer $(2(R)-4g, R = CH_2CH_2CO_2CH_3)$ via Epimerization. The protocol for carrying out the epimerization was the same as that described above for 4a, (R = Me); the reaction time was 5 h.

(3(2S),4S)-3-(2-(N,N'-bis-(t-butoxycarbonyl)hydrazino)-3-methyl-1-oxo-3-butenyl)-4-(phenyl-methyl)-2-oxazolidimone (4h, R = C(=CH₂)Me and B- and Z-(4S)-3-(4-(N,N'-bis-(t-butoxy-carbonyl)hydrazino)-3-methyl-1-oxo-2-butenyl)-4-(phenylmethyl)-2-oxazolidimone (7). Following an exact scale up of the general procedure, 2.59 g (10.0 mmol) of 4h gave 2.25 g (47%) of 2S (R = C(CH₃)=CH₂) as a colorless glass foam after purification by MPLC (650 g of silica gel; 6 L of Solvent A, 2 L linear gradient from Solvent A to CH₂Cl₂-bexane-CH₃CN (70: 30: 15), and finally 2 L of CH₂Cl₂-bexane-CH₃CN (70: 30: 15)). HPLC analysis (vide supra, Solvent A) of the unchromatographed product gave a 2(S) (t_T = 5.87 min): 2(R) (t_T = 7.50 min) ratio of 98: 2. HPLC analysis of the purified product gave a 2(S): 2(R) ratio of >300: 1: IR (CH₂Cl₂) 3405, 1785, 1748, 1695 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 331 °K) δ 7.36-7.18 (m, 5H, aromatics), 6.64 (very broad s, 1H, NH), 6.262 (s, 1H, CHC=O), 5.019 (s, 1H, HHC=C), 4.696 (s, 1H, HHC=C), 4.70-4.56 (sym m, 1H, CH₂Cl₂N), 4.20-4.13 (m, 2H, OCH₂), 3.298 (dd, J_{ViC} = 3.0 Hz, J_{gem} = 13.7 Hz, 1H, CH₂HC₆H₅), 2.827 (dd, J_{ViC} = 9.3 Hz, J_{gem} = 13.6 Hz, 1H, CH₂C₆H₅), 2.002 (s, 3H, CH₃), 1.489 (s) and 1.445 (s) (18H, OC(CH₃)₃); { α ₁D₂²⁶ +209° (c = 1.04, CHCl₃). Anal. Calcd for C₂SH₃SN₃O₇: C, 61.33; H, 7.21. Found: C, 61.48; H, 7.13.

After several later fractions (149 mg, 3.0%) found by HPLC to be a 95: 5 mixture of 2(S) and 2(R) diastereomers, respectively, there was obtained 52 mg (1.1%) found by HPLC to be a 29: 71 mixture of 2(S)-4h and 2(R)-4h, respectively. H NMR (250 MHz, CDCl₃, 331 °K) integration of this material gave a 2(S): 2(R) ratio of 30: 70. Distinctive signals attributed to the minor diastereomer (2(R)-4h) are: $8 \cdot 6.208$ (br s, CHC=0), 5.091 (s, HHC=C), 4.758 (s, HHC=C), 3.416 (dd, I_{ViC} = 3.4 Hz, I_{Rem} = 13.2 Hz, CHHC₆H₅), 2.036 (s, CH₃), 1.472 (s, OC(CH₃)₃).

Still later fractions yielded 2.24 g (42%) of the strongly UV absorbing 1,4-addition product 7 as a colorless glass foam. This material was found by HPLC and $^1\mathrm{H}$ NMR to be a 6: 4 mixture of geometrical (E,Z) isomers: HPLC (Solvent B) $t_T=10.4$, 11.8 min; TLC R_f 0.20 (silica, Solvent B); IR (CHCl3) 3395 (br), 3063, 2980, 2935, 1780, 1745, 1713, 1685, 1643, 1480, 1455, 1393, 1385, 1368, 1350, 1173, 1152 cm⁻¹; $^1\mathrm{H}$ NMR (300 MHz, CDCl3, 330 °K) & 7.34-7.17 (m, 5H, C6H5), 6.978 (q, J = 1.3 Hz, 0.4H, C=CH in minor isomer), 6.908 (q, J = 1.3 Hz, 0.4H, C=CH in major isomer), 4.76-4.65 (m, 1.8H, C=CH in minor isomer and CH2CHN), 4.26-4.09 (m, 3.2H, CH2NCO2 in major isomer and CH2O), 3.356 (dd, J = 3.4, 13.4 Hz) and 3.319 (dd, J = 3.4, 13.4 Hz) (1H total, CHHC6H5), 2.780 (dd, J = 9.5, 13.4 Hz) and 2.772 (dd, J = 9.5, 13.4 Hz) (1H total, CHHC6H5), 2.150 (d, J = 1.0 Hz, 1.8H, CH3 in major isomer), 2.018 (d, J = 1.3 Hz, 1.2H, CH3 in minor isomer), 1.478 (s), 1.472 (s), and 1.464 (s) (18H total, OC(CH3)3). Anal. Calcd for C25H35N3O7: C, 61.33; H, 7.21. Found: C, 61.35; H, 7.34.

General Procedure for the Magnesium Methoxide Methanolysis of Carboximides 4 To 4.0 mL of dry MeOH, cooled to 0 °C under N2, was added dropwise 1.00 mmol of MeMgBr (0.315 mL of a 3.2 M solution in Et2O). After stirring the above solution at 0 °C for 5 min, a solution of 0.50 mmol of the imide, 2, in 4.0 mL in dry MeOH was added via cannula, residual 2 being rinsed in with two 2 mL-portions of dry MeOH. The above solution was stirred at 0 °C for an appropriate time (30-90 min, vide infra) and then quenched with 4 mL of pH 7 phosphate buffer. The mixture was partitioned between half-saturated aqueous NaCl/NH4Cl and CH2Cl2. The aqueous phase was washed with CH2Cl2 (x 3). The CH2Cl2 phases were combined, dried (MgSO4), and evaporated in vacuo. The residual unpurified product was purified by MPLC.

General Procedure for the Lithium Hydroxide Hydrolysis and Dinzomethane Esterification of Carboximides 4. An ice cooled solution of 0.50 mmol of the imide, 2, in 2.0 mL of THF was treated in one portion with a cold (0 °C) solution of 28 mg (1.2 mmol; 2.3 equiv) of LiOH in 1.0 mL of H₂O. The resulting two-phase mixture was stirred at 0 °C until the reaction was complete (vide infra) and was then worked up by one of the following methods:

Method A: The mixture was partitioned between CH₂Cl₂ (20 mL) and H₂O (20-30 mL) containing sufficient NaCl to break the emulsion. The aqueous phase was washed with CH₂Cl₂ (3 x 20 mL). The CH₂Cl₂ phases were combined, dried (MgSO₄), and evaporated in vacuo. Purification of the residue by MPLC afforded the chiral auxiliary, XpH. The aqueous phase was acidified with 1 N aqueous NaHSO₄ and extracted with CH₂Cl₂ (4 x 30 mL). The CH₂Cl₂ extracts were combined, dried (Na₂SO₄), and concentrated in vacuo. A CH₂Cl₂ solution of the residual unpurified acid was treated dropwise at 0 °C with ethereal diazomethane until a yellow color persisted. Following a standard workup (vide infra), the methyl ester was purified by MPLC.

Method B: The mixture was partitioned between 1 N aqueous NaHSO₄ and CH₂Cl₂. The aqueous phase was extracted with CH₂Cl₂ (x 3), and the organic phases were combined, dried (Na₂SO₄), and evaporated in vacuo. A solution of the residue in CH₂Cl₂ (20 mL) was treated dropwise at 0 °C with ethereal CH₂N₂ until a yellow color persisted. The solution was decolorized with HOAc, washed with saturated aqueous NaHCO₃, dried (MgSO₄) and evaporated in vacuo. The product was purified by MPLC.

 $2S \cdot (N,N'-Bis \cdot (t-butoxycarbonyl)hydraxino)-3-phenyl-1-propanoic Acid, Methyl Ester (9c, R <math>\pm$ CH₂Ph). Method A (Table 3, Entry A). Lithium Hydroxide hydrolysis (0 °C, 3 h; workup b) of 270 mg (0.500

mmol of 4c (R = CH₂Ph) afforded, after CH₂N₂ treatment, 161 mg (82%) of 9c (R = CH₂Ph) after purification by flash chromatography (10 g of silica gel; hexane-EtOAc (3:1)). The optical purity of this material was found by capillary gas chromatographic analysis of its derived (+)-MTPA amide, 10c (R = CH₂Ph), to be >99% ee (vide infra). The chiral suxiliary, XpH, was recovered in 85% yield by MPLC (50 g of silica gel; hexane-EtOAc (1:1)).

Method B (Table 3, Entry B). Magnesium methoxide methanolysis (0 °C, 30 min) of 270 mg (0.500 mmol) of 4c (R = CH₂Ph) afforded 176 mg (89%) of 9c (R = CH₂Ph) as a colorless, viacous oil after purification by MPLC (50 mg of silica gel; 2L linear gradient from hexane-EtOAc (9:1) to hexane-EtOAc (1:1)): IR (CHCl₃) 3395, 1743, 1713 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 330 °K) δ 7.27-7.16 (m, 5H, aromatics), 6.240 (br s, 1H, NH), 5.00-4.95 (unresolved broad m, 1H, CHC=O), 3.656 (s, 3H, OCH₃), 3.169 (d, J = 7.2 Hz, 2H, CHC=O), 1.434 (s) and 1.421 (s) (18 H, OC(CH₃)₃); $\{\alpha\}_D^{22}$ -19.9° (c = 1.21, CHCl₃). Anal. Calcd for C₂OH₃ON₂O₆: C, 60.90; H, 7.67. Found: C, 60.84; H, 7.76. The optical purity of the above material was found by capillary gas chromatographic analysis of its derived (+)-MTPA amide, 10e (R, = CH₂Ph), to be >99% ee (vide infra). Later fractions afforded 176 mg (89% recovery) of the chiral auxiliary, XpH.

2S-(N,N'-Bis-(t-butoxycarbonyl)hydrazino)phenylacetic Acid, Methyl Ester (9d, R = Ph) Method A.(Table 3, Entry D) Lithium hydroxide hydrolysis (0 °C, 120 min; workup b) of 263 mg (0.50 mmol) of 4d (R = Ph) afforded, after CH₂N₂ treatment, 160 mg (84%) of 9d (R = Ph) after purification by MPLC (50 g of silica gel; hexane-EtOAc (4:1)): IR (CHCl₃) 3410, 1748, 1707 cm⁻¹; ¹H NMR (250 MHz; CDCL₃; 331 °K) δ 7.306 (s, 5H, aromatics), 6.40 (very broad s, 1H, NH), 5.95 (br s, 1H, CHC=O), 3.742 (s, 2H, OCH₂), 1.478 (s, 9H, OC(CH₃)₃), 1.193 (br s, 9H, OC(CH₃); α ₁|₀²² +151.6° (c = 1.96, CHCl₃). Anal. Calcd for C₁9H₂8N₂O₆: C, 59.89; H, 7.42. Found: C, 60.17; H, 7.51. The optical purity of this material was found by capillary GLC analysis of its derived (+)-MTPA-amide, 10d, to be 98% ee.

Method B (Table 3, Entry E). Magnesium methoxide methanolysis (0 °C, 90 min) of 263 mg (0.50 mmol) of 4d (R = Ph) afforded 140 mg (71%) of 3d (R = Ph) after purification by MPLC. The optical purity of this material was found by capillary gas chromatographic analysis of its derived (+)-MPTA-amide, 18d (R = Ph), to be 93% ee (vide infra).

General Procedure for the Lithium Benzyloxide Transesterification ¹⁹ of Carboximides 4. To a cold (-78 °C) solution of 2.00 mmol of LiOBn (prepared as previously described from 310 µL (324 mg; 3.00 mmol) of redistilled benzyl alcohol and 2.00 mmol of n-butyl lithium (1.59 M in hexane)) in 6.0 mL of dry THF was added via cannula a precooled (-78 °C) solution of 1.00 mmol of the imide 2 in 4.0 mL of THF. Residual imide was rinsed in with two 2 mL-portions of THF, and the resulting solution stirred at -50 °C for 2.0-50 h (vide infra) prior to quenching with 6 mL of aqueous pH 7 phosphate buffer. The mixture was partitioned between H₂O and CH₂Cl₂. The aqueous phase was extracted with three portions of CH₂Cl₂ and the organic phases were combined, dried (MgSO₄) and evaporated in vacuo. The unpurified product was purified by MPLC:

2S-(N,N'-Bis-(t-butoxycarbonyl)hydraxino)-3-phenylpropanoic Acid, Benzyl Ester (6c, R = CH₂Ph) (Table 3, Entry C). As described above, lithium benzyloxide transesterification (-50 °C, 2.0 h) of 540 mg (1.00 mmol) of 4c (R = CH₂Ph) afforded 451 mg (96%) of 6c (R = CH₂Ph) as a colorless glass after purification by MPLC (50 g of silica gel; 0.5 L of hexane-EtOAc (9:1), 1.0 L linear gradient from hexane-EtOAc (9:1) to EtOAc, and finally EtOAc): IR (CHCl₃) 3395, 1743, 1713 cm⁻¹; ¹H NMR (250 MHz, CDCl₃, 331 °K) δ 7.33-7.14 (m, 10H, aromatics), 6.226 (br s, 1H, NH), 5.088 (s, 2H, CHCH₂Ph), 5.07-5.00 (unresolved m, 1H, CHC=O), 3.185 (d, J = 7.2 Hz, OCH₂Ph), 1.420 (s) and 1.399 (s) (18H, OC(CH₃)₃); [α]_D²² -4.16° (c = 1.99, CHCl₃). Anal. Calcd for C₂6H₃4N₂O₆: C, 66.36; H, 7.28. Found: C, 66.41; H, 7.18. The optical purity of the this material was found to be >99% ee by capillary GLC analysis of its derived (+)-MPTA-amide, 10c (R = CH₂Ph) (vide infra). Later fractions afforded 171 mg (96% recovery) of the

2S-(N,N'-Bis-(t-butoxycarbonyl)hydrazino)-3-methylbutanoic Acid, Benzyl Ester (6e, R = i-Pr) (Table 3, Entry H). In an exact scale up of the general lithium benzyloxide transesterification procedure (-50 °C, 15.5 h), 1.97 g (4.00 mmol) of 4e (R = i-Pr) gave 1.38 g (82%) of 6e (R = i-Pr) as a viscous oil after purification by MPLC (165 g of silica gel; 1 L of hexane-EtOAc (90:10), 1 L of hexane-EtOAc (75:25) and finally 1 L of EtOAc): IR (Neat) 3335, 1742, 1713 cm⁻¹; ¹H NMR (250 MHz, CDCl₃, 331 °K) & 7.35-7.27 (m, 5H, aromatics), 6.354 (br s, 1H, NH), 5.152 (s, 2H, OCH₂), 4.534 (br d, J = 6.6 Hz, 1H, CHC=O), 2.36-2.22 (sym 6 line m, 1H, CH(CH₃)₂), 1.456 (s) and 1.431 (s) (18H, OC(CH₃)₃), 1.048 (d, J = 6.8, 3H, CHCH₃), 1.023 (d, J = 6.8 Hz, 3H, CHCH₃); $(\alpha)_D^{22}$ -7.03° (c = 1.89, CHCl₃). Anal. Calcd for C₂₂H₃₄N₂O₆: C, 62.54; H, 8.11. Found: C, 62.70; H, 8.12. The optical purity of the above material was found to be >99% ee by capillary gas chromatographic analysis of its derived (+)-MTPA-amide 10e (vide infra).

2(S)-(N,N'-bis(t-butoxycarbonyl)hydrazino)-3,3-dimethylbutamoic acid, methyl ester (9f, R = t-Bu). Method A.(Table 3, Entry K). A solution of 253 mg (0.500 mmol) of 4f in 7.5 mL of THF and 2.3 mL of H₂O, stirred at 0 °C under N₂, was treated with 0.20 mL (2.0 mmol, 4.0 equiv) of 31% H₂O₂ followed by 24 mg (1.0 mmol, 2.0 equiv) of LiOH (anhydrous powder). The resulting mixture was stirred at 0 °C for 3.25 h, and was then treated

with a solution of 280 mg (2.2 mmol) of Na₂SO₃ in 1.5 mL of H₂O. Following the addition of 5 mL of 0.5 N aqueous NaHCO₃, the THP was evaporated in vacuo. The aqueous residue was acidified with 1 N aqueous NaHSO₄ and extracted with four portions of CH₂Cl₂. The CH₂Cl₂ extracts were combined, dried (Na₂SO₄), and concentrated in vacuo. The resulting solution (15 mL) was treated dropwise at 0 °C with ethereal diazomethane until a yellow color persisted. The solution was decolorized with glacial HOAc, washed with saturated aqueous NaHCO₃, dried (MgSO₄), and evaporated in vacuo. The residue was chromatographed on one size B Michel-Miller column (50 g of silica gel) eluting with 0.5 L of hexane-EtOAc (85: 15) followed by a 1L linear gradient from hexane-EtOAc (60: 40) to EtOAc. Early fractions yielded 165 mg (91%) of the ester, 9f, as a white solid: IR (CHCl₃) 3385 (br), 1745, 1707 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 331 °K) & 6.45 (br s, 1H, NH), 4.658 (s, 1H, CHC=O), 3.707 (s, 3H, OCH₃), 1.468 (s) and 1.462 (s) (18H, OC(CH₃)₃). 1.124 (s, 9H, CHC(CH₃)₃); $\{\alpha_i\}_D^{23}$ -15.4° (c = 1.03, CHCl₃); mp 76.5-77.5 °C. Anal. Calcd for C₁7H₃2N₂O₆: C, 56.65; H, 8.95. Found: C, 56.68; H, 9.01. The optical purity of the above material was found to be >99% ee by capillary gas chromatographic analysis of its derived (+)-MTPA-amide 10f (vide infra). Later fractions afforded 14.6 mg (6.1%yield) of the β-hydroxyethyl amide ring opened product followed by 80.6 mg (91% recovery) of the chiral auxiliary, 2 (XpH).

Method B (Table 3, Emtry I). This is a modification of the general LiOH hydrolysis procedure. A solution of 253 mg (0.500 mmol) of 4f in 7.5 mL of THF and 2.5 mL of H₂O, stirred at 0 °C under N₂, was treated with 24 mg (1.0 mmol, 2.0 equiv) of LiOH (anhydrous powder). The resulting mixture was stirred at 0 °C for 16 h. Following the addition of 5 mL of 0.5 N aqueous NaHCO₃, the THF was evaporated in vacuo. The aqueous residue was acidified with 1 N aqueous NaHSO₄ and extracted with four portions of C₂HCl₂. The CH₂Cl₂ extracts were combined, dried (Na₂SO₄), and concentrated in vacuo. The resulting solution (15 mL) was treated dropwise at 0 °C with ethereal diazomethane until a yellow color persisted. The solution was decolorized with glacial HOAc, washed with saturated aqueous NaHCO₃, dried (MgSO₄), and evaporated in vacuo. The residue was chromatographed (MPLC) on one size B Michel-Miller column (50 g of silica gel) eluting with 0.5 L of hexane-EtOAc (90: 10) followed by a 1L linear gradient from hexane-EtOAc (60: 40) to EtOAc. Early fractions yielded 29.6 mg (16%) of the ester, 9f, as a white solid. The optical purity of the above material was found to be >99% ee by capillary gas chromatographic analysis of its derived (+)-MTPA-amide 10f (vide infra). Later fractions afforded 185.3 mg (76%yield) of the hydroxyethyl amide ring opened product,

2S-(N,N'-Bis-(t-butoxycarbonyl)hydrazino)-3,3-dimethylbutanoic Acid, Benxyl Ester (6f, R = t-Bu) (Table 3, Entry J). In an exact scale up of the general lithium benzyloxide transesterification procedure (-50 °C, 50 h), 2.75 g (5.44 mmol) of 4f (R = t-Bu) gave 1.21 g (51%) of 6f (R = t-Bu) as a viscous oil after purification by MPLC (in two portions on 165 g of silica gel; hexane-EtOAc (93:7)): IR (CHCl3) 3490, 1747, 1708 cm⁻¹; ¹H NMR (250 MHz, CDCl3, 331 °K) δ 7.35-7.28 (m, 5H, aromatics), 6.39 (very br s, 1H, NH), 5.160 (s, 2H, OCH2), 4.701 (br s, 1H, CHC=O), 1.454 (s) and 1.434 (s) (18H, OC(CH3)3), 1.119 (s, 9H, CHC(CH3)3); $[\alpha]_D^{22}$ -8.97° (c = 2.75, CHCl3). Anal. Calcd for C23H36N2O6: C, 63.28; H, 8.31. Found: C, 63.38; H, 8.19. The optical purity of the this material was found to be >99% ee by capillary gas chromatographic analysis of its derived (+) MTPA-amide, 10f (R = t-Bu) (vide infra).

General Procedure for the Deprotection, Hydrogenolysis and Acylation of the N,N'-Bis(t-butoxycarbonyl)- α -hydrazino Methyl Esters, 9: Assay for Optical Purity. A stirred solution of 0.100 mmol of the protected hydrazino ester 9 in 2.0 mL of CH₂Cl₂ was treated with 2.0 mL of trifluoroacetic acid (TFA) at 25 °C under N₂. After 30 min, the Raney Nickel³⁸ was added and the stirred mixture hydrogenated under 550 PSIG H₂ for 4-16 h (vide infra). The mixture was purged with N₂ and then filtered through Celite. The filter cake was washed with MeOH (x 4) and the filtrate evaporated in vacuo, residual TFA and MeOH being removed by azeotroping with toluene (x 3). The glassy green residue containing the α -amino ester was suspended in CH₂Cl₂ (3 mL/0.10 mmol) and acylated by treatment with 2.0 equiv of (+) MTPA-chloride ³³ and 4.0 equiv of triethylamine at 25 °C. After stirring for 3 h at 25 °C, 2 mL of H₂O was added and stirring was resumed for 1 h additional. The mixture was diluted with CH₂Cl₂ and washed with 1 N HCl. The aqueous phase was back-extracted with CH₂Cl₂ (x 2). The CH₂Cl₂ phases were combined, washed with saturated aqueous NaHCO₃, dried (MgSO₄), and evaporated in vacuo. The residue was purified by careful flash chromatography such that no fractionation of diastereomers was effected. The diastereomeric purity of the resulting (+)-MTPA-amide, 10, was determined by capillary GLC analysis on a 0.25 mm x 30 m DB-1 fused silica column (vide infra):

2S-((+)- α -Methoxy- α -trifluoromethylphenylacetylamino)-3-phenylpropionic Acid, Methyl Ester, 10c (R = CH₂Ph) (Table 4, Entry A). A sample (47.9 mg; 0.100 mmol) of 6c (R = CH₂Ph), secured via LiOBn transesterification (vide supra), was converted to the corresponding methyl ester 9c (R = CH₂Ph) by debenzylation (1 atm of H₂, 7 mg of 5% Pd/C, 1.5 mL of EtOAc, 3 h, 25 °C) and subsequent diazomethane treatment (CH₂Cl₂, 0 °C). Without purification, 9c was subjected to the above deprotection, hydrogenolysis (4 h), and acylation ((+)-MTPA-chloride) ³³ sequence. Purification by flash chromatography (6 g of silica gel; hexane-EtOAc (4:1)) afforded 37.7 mg (94%) of 10c (R = CH₂Ph) as a white solid: IR (CHCl₃) 3415, 1747, 1698 cm⁻¹; ¹H NMR (250 MHz, CDCl₃) δ 7.52-7.13 (m, 11H, aromatics and NH), 4.911 (ddd, J = 5.7, 6.7, 8.0 Hz, 1H, CHC=O), 3.734 (s, 3H, CO₂CH₃), 3.73-3.08 (AB m

and q at 3.215 (J_{HF} = 1.5 Hz), 5H, CH₂Ph and CF₃COCH₃); ¹³C NMR (62.5 MHz, CDCl₃) δ 171.34, 165.96, 135.54, 132.09, 129.45, 129.15, 128.65, 128.45, 127.93, 127.24, 123.73 (q, J_{CF} = 290.2 Hz), 84.12 (q, J_{CF} = 26.1 Hz), 54.77, 53.16, 52.29, 37.72. Capillary GLC analysis (190 °C, 15 PSI) gave a 2(S)-10c (t_r = 7.04 min): 2(R)-10c (t_r = 6.64 min) ratio of 230:1. The above solid was recrystallized from acetone-hexane: mp 99.5-101 °C; $[\alpha]_D^{2.5}$ +6.77° (c = 1.00, CHCl₃). Anal. Calcd for C₂₀H₂₀F₃NO₄: C, 60.75; H, 5.10. Found: C, 60.82; H, 5.22.

A sample of L-phenylalanine methyl ester hydrochloride (10.8 mg; 0.050 mmol) in CH₂Cl₂ (0.5 mL) was treated as described above with (+)-MTPA-chloride 33 (14.5 μ L, 0.075 mmol, 1.5 equiv) and Et₃N (21 μ L, 0.15 mmol, 3.0 equiv) for 2 h at 25 °C. Purification by flash chromatography afforded 18.6 mg (94%) of authentic 10c (R = CH₂Ph), found by 1 H NMR and capillary GLC coinjection to be identical to the major diastereomer in the imide derived sample.

Acylation of L-phenylalanine methyl ester hydrochloride with (-)-MTPA-chloride³³ gave, after purification by flash chromotography, 19.0 mg (96%) of the (-)-MTPA-amide ($R = CH_2Ph$) as a colorless oil: ¹H NMR (250 MHz; CDCl₃) & 7.40-7.13 (m, 8H, aromatics), 7.015 (br d, J = 8.3 Hz, 1H, NH), 6.91-6.87 (m, 2H, aromatics), 4.991 (ddd, J = 5.4, 7.0, 8.4 Hz, 1H, CHC=O), 3.757 (s, 3H, CO₂CH₃), 3.431 (q, $J_{HF} = 1.6$ Hz, 3H, CF₃COCH₃), 3.185-2.983 (8 line AB portion of ABX system, $J_{AB} = 14.0$ Hz, $J_{AX} = 7.0$ Hz, $J_{BX} = 5.4$ Hz, 2H, CH₂Ph). This material was shown by coinjection to have the same retention time on capillary GLC as the minor diastereomer, 2(R)-10c ($R = CH_2Ph$), in the imide derived sample.

2S-((+)- α -Methoxy- α -trifluoromethylphenylacetylamino)phenylacetic Acid, Methyl Ester, 10d (R = Ph) (Table 4, Entry B). A sample (38.4 mg; 0.10 mmol) of 9d (R = Ph), secured via LiOH-hydrolysis/CH₂N₂-esterification (vide supra), was subjected to the above deprotection, hydrogenolysis (4 h) sequence. The impurified amino ester was divided into two equal portions as a MeOH solution. Following solvent removal (in vacuo, tohiene azeotrope) one portion was acylated as described above by treatment with 19.4 μ L (0.10 mmol; 2.0 equiv) of (+)-MTPA-chloride³³ and 28 μ L (0.20 mmol, 4.0 equiv) of triethylamine in 1.5 mL of CH₂Cl₂. The unpurified product was purified by flash chromatography (4 g of silica gel; hexane-EtOAc (85:15)) to afford 19.0 mg (99%) of 10d (R = Ph) as a colorless, viscous oil: IR (CHCl₃) 3415, 1748, 1700 cm⁻¹; ¹H NMR (250 MHz; CDCl₃) δ 7.771 (br d, J = 6.7 Hz, 1H, NH), 7.60-7.57 (m, 2H, aromatics), 7.45-7.34 (m, 8H, aromatics), 5.584 (d, J = 7.2 Hz, 1H, CHC=O), 3.733 (s, 3H, CO₂CH₃), 3.358 (q, J_{HF} = 1.3 Hz, CF₃COCH₃); ¹³C NMR (62.5 MHz, CDCl₃) δ 170.68, 165.85, 135.68, 131.94, 129.54, 129.07, 128.77, 128.57, 128.03, 127.28, 123.76 (q, J_{CF} = 290.3 Hz), 84.14 (q, J_{CF} = 25.7 Hz), 56.60, 54.88, 52.78; [α]_D²² +80.9° (c = 0.90, CHCl₃); High Resolution MS. Calcd for C₁9H₁₈F₃NO₄: m/e 381.11878. Found: m/e 381.11857. Capillary GLC diastereomer analysis (200 °C, 5 PSI) gave a 2(S)-10d (t_f = 10.65 min): 2(R)-10d (t_f = 9.8 min) ratio of 99:1.

The second portion of unpurified amino ester (vide supra) was acylated exactly as described above with (-)-MTPA-chloride 33 to afford 19.1 mg (99%) of the corresponding (-)-MTPA-amide (R = Ph) as a colorless glass: 1 H NMR (250 MHz; CDCl₃) 5 7.610 (br d, J = 6.5 Hz, 1H, NH), 7.41-7.25 (m, 10H, aromatics), 5.605 (d, J = 7.3 Hz, 1H, CHC=O), 3.753 (s, 1H, CO₂CH₃), 3.549 (q, J_{HF} = 1.6 Hz, 3H, CF₃COCH₃). This material was shown by coinjection with the above sample of 10d (R = Ph) to have the same retention time on capillary GLC as the minor diastereomer, 2 (R)-10d, in that sample.

2S-((+)- α -Methoxy- α -trifluoromethylphenylacetylamino)-3-methylbutamoic Acid, Methyl Ester, 10e (R = *l*-Pr) (Table 4, Entry C). A sample (135 mg; 0.319 mmol) of 6e (R = *i*-Pr), secured via LiOBn transesterification (vide supra) was converted to the corresponding methyl ester 9e by debenzylation (1 atm of H₂, 17 mg of 5% Pd/C, 5 mL of EtOAc, 3 h, 25 °C) and subsequent diazomethane treatment (CH₂Cl₂, 0 °C). Without purification, 9e was subjected to the above deprotection (4 mL of CH₂Cl₂, 4 mL of TFA), hydrogenolysis (16 h), and acylation ((+)-MTPA-chloride)³³ sequence. Purification by flash chromatography (35 g of silica gel; hexane-EtOAc (85:15)) afforded 92.0 mg (83%) of 10e (R = *i*-Pr) as a pale yellow oil: IR (Neat) 3430, 3350 (sh), 1746, 1705 cm⁻¹; ¹H NMR (250 MHz, CDCl₃) δ 7.56-7.52 (m, 2H, aromatic), 7.46-7.37 (m, 4H, aromatic and NH), 4.597 (dd, J = 4.7, 9.1 Hz, 1H, CHC=O), 3.743 (s, 3H, CO₂CH₃), 3.387 (q, J_{HF} = 1.3 Hz, 3H, CF₃COCH₃), 2.281 (d of septets, J = 4.7, 6.9 Hz, 1H, CH(CH₃)), 0.994 (d, J = 6.9 Hz, 3H, CH(CH₃)), 0.947 (d, J = 6.9 Hz, 3H, CH(CH₃)), 13.05 (q, J_{CF} = 290.2 Hz), 84.29 (q, J_{CF} = 25.9 Hz), 57.25, 54.83, 52.01, 31.14, 18.90, 17.59; [α]_D²⁵-23.8° (c = 0.92, CHCl₃). High Resolution MS. Calcd for C₁6H₂0F₃NO₄: m/e 347.13443. Found: m/e 347.13369. Capillary GLC diastereomer ysis (150 °C, 7 PSI) gave a 2(S)-10e (t_T = 13.59 min): 2(R)-10e (t_T = 13.03 min) ratio of >500:1.

A sample of L-valine methyl ester hydrochloride (16.8 mg, 0.10 mmol) in CH₂Cl₂ (1.0 mL) was treated as described above with (+)-MTPA-Chloride³³ (29.1 μ L, 37.9 mg, 0.15 mmol) and Et₃N (41.8 μ L, 30.4 mg, 0.30 mmol) for 3 h at 25 °C. Purification by flash chromatography afforded 34.9 mg (100%) of 19e (R = i-Pr) found by ¹H NMR and capillary GLC coinjection to be identical to the major diastereomer in the imide derived sample.

Acylation of L-valine methyl ester hydrochloride with (-)-MTPA-chloride³³ in an identical fashion gave, after purification by flash chromatography, 32.8 mg (95%) of the corresponding (-)-MTPA-amide (R = i-Pr) as a colorless oil: ¹H

NMR (250 MHz, CDCl₃) δ 7.60-7.56 (m, 2H, aromatic), 7.45-7.37 (m, 3H, aromatic), 7.051 (br d, J = 9.0 Hz, 1H, NH), 4.617 (dd, J = 4.6, 9.1 Hz, 1H, CHC=O), 3.764 (s, 3H, CO₂CH₃), 3.553 (q, $J_{HF} = 1.7$ Hz, 3H, CP₃COCH₃), 2.200 (d of septets, J = 4.7, 6.9 Hz, 1H, CH(CH₃)₂), 0.871 (d, J = 6.9 Hz, 3H, CH(CH₃)), 0.808 (d, J = 6.9 Hz, 3H, CH(CH₃)). This material was shown by coinjection to have the same retention time on capillary GLC as the minor diastereomer, 2(R)-19e (R = i-Pr), in the imide derived sample.

2S-((+)- α -Methoxy- α -trifluoromethylphenylacetylamino)-3,3-dimethylbutawoic Acid, Methyl Ester, 10f (R = t-Bu) (Table 4, Entry D). A sample (43.6 mg; 0.100 mmol) of 6f (R = t-Bu), secured via LiOBn transesterification (vide supra), was converted to the corresponding methyl ester 9f by debenzylation (1 atm of H₂, 7 mg of 5% Pd/C, 1.5 mL of EtOAc, 3 h, 25 °C) and subsequent diazomethane treatment (CH₂Cl₂, 0 °C). Without purification, 9f was subjected to the above deprotection, hydrogenolysis (16 h) sequence. The unpurified amino ester was divided into two equal portions as a MeOH solution. Following solvent removal (in vacuo, toluene azeotrope), one portion was

of silica gel; hexane-EtOAc (9:1)) to afford 16.0 mg (89%) of 18f (R = t-Bu) as a colorless viscous oil: IR (neat) 3430, 1744, 1708 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.52-7.48 (m, 3H, aromatic and NH), 7.43-7.38 (m, 3H, aromatic), 4.463 (d, J = 9.5 Hz, 1H, CHC=O), 3.727 (s, 3H, CO₂CH₃), 3.390 (q, J_{HF} = 1.4 Hz, 3H, CP₃COCH₃), 1.025 (s, 9H, C(CH₃)₃); ¹³C NMR (62.5 MHz; CDCl₃) δ 171.40, 165.86, 131.98, 129.45, 128.54, 128.05, 123.91 (q, J_{CF} = 290.1 Hz) 84.32 (q, J_{CF} = 25.9 Hz), 60.39, 54.95, 51.77, 34.87, 26.64; $\left[\alpha\right]_D^{25}$ -28.8° (c = 0.71, CHCl₃). High Resolution MS. Calcd for C₁₇H₂₂F₃NO₄: m/e 361.15008. Found: m/e 361.14596. Capillary GLC analysis (150 °C, 6 PSI) gave a 2(S)-10f (t_r = 17.00 min): 2(R)-10f (t_r = 16.31 min) ratio >500:1.

The second portion of unpurified amino ester (vide supra) was acylated exactly as described above with (-)-MTPA-chloride.³³ The unpurified product was purified as described above by flash chromatography to afford 15.2 mg (84%) of the corresponding (-)-MTPA-amide (R = t-Bu) as a colorless viscous oil: ¹H NMR (300 MHz, CDCl₃) δ 7.60-7.57 (m, 2H, aromatic), 7.43-7.38 (m, 3H, aromatic), 7.116 (br d, J = 9.7 Hz, 1H, NH), 4.483 (d, J = 9.7 Hz, 1H, CHC=O), 3.755 (s. 3H, CO₂CH₃), 0.903 (s, 9H, C(CH₃)₃). This material was shown by coinjection with 10f (R = t-Bu) to have the same retention time on capillary GLC as the minor diastereomer, 2(R)-10f (R = t-Bu), in that sample.

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References and Notes

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